

NGS + dPCR for Hematologic Malignancies

From comprehensive molecular profiling to highly sensitive, quantitative follow-up testing of known mutations and fusion targets.

Two Partners. one hematology solution

Kindstar Global

Founded in 2003, Kindstar is Asia's leading independent medical laboratory group (Stock code: 9960.HK), offering patients and physicians access to advanced esoteric testing across hematology, gynecology and pediatrics, infectious diseases, oncology and neurology.

Hematology is at the core of its work. Kindstar uses NGS to find molecular targets and RainSure dPCR to follow them over time.



25+ years

Clinical experience in hematologic diseases

~6,000

Patients tested per month

3,000+

Hospitals covered
including over 1,000 tertiary hospitals

Top 1

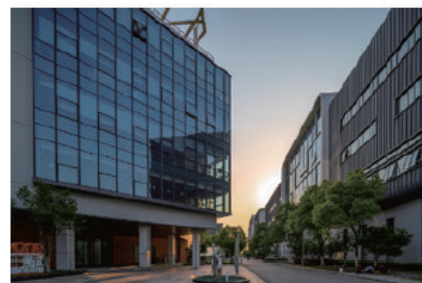
Asia's leading specialty testing provider in hematologic diseases

RainSure Scientific

Founded in 2017, RainSure Scientific is a digital PCR company that designs, develops and manufactures its own droplet dPCR systems, reagents and assays, with R&D in San Jose, California, and manufacturing in Suzhou, China.

From instruments to assays, every solution is engineered for performance, scalability, and trust. Our portfolio includes CE-IVD, FDA 510(k) and RUO offerings, produced at manufacturing sites operating under ISO 13485 and certified medical device standards.

RainSure droplet dPCR delivers absolute quantification without standard curves, in a sealed workflow that reduces contamination risk. This makes it ideal for repeated, low-level MRD monitoring in hematology.



9 years

Innovating digital PCR product development

50+

Patents and IP

CE-IVD / FDA

Authoritative certifications

4,000 m²

ISO 13485 GMP facility

A laboratory perspective

"With thousands of patient samples tested every month, we need a digital PCR platform that is reliable, scalable, and easy for our team to operate. RainSure digital PCR has supported our high-throughput testing workflow with consistent performance and operational efficiency."

Feng Wang, PhD

Chief Scientist, Hematology Diagnostic Service Business Unit, Kindstar Global

RainSure dPCR platforms

DropDx System



RS32 System



Shared features – both DropDx and RS32

Shared features

MonoFlex™ droplet technology: uniform droplets, size CV $\leq 3\%$

Absolute quantification: Poisson statistics, no standard curve

Dynamic range: 5 logs

Time to results: 40 cycles < 90 mins*

Sensitivity: 0.001% by merging 4 wells*

Sealed workflow: no opening after loading, reduced aerosol contamination

Flat-Fast™ flat heating: fast, uniform PCR on a flat surface; gradient PCR supported

Universal Macro™ reagent platform: compatible with third-party qPCR reagents, intercalating dye and probe-based chemistries

Consumable: Cartridge, droplet oil, Mastermix, Cartridge Seal, Heat film

Image-based reading: fluorescence images for result review

Raw droplet data: Accessible for data verification

Sample recovery / re-testable results: PCR products can be retrieved from cartridges applying an additional fixture; cartridges can be stored and re-scanned

Different features

Features	DropDx	RS32
Instrument name	DropDx (CE-IVD model) DropMDx (FDA 510(k) model)	RS3204/RS3205/RS3206/RS3207
Configuration	Integrated droplet generation and PCR unit, with a strategically separated high-speed reader	All-in-one: droplet generation, PCR and reading in one instrument
Workflow and scale	Several generation/PCR units can run in parallel with staggered loading; one reader serves multiple units	Fully automated after cartridge loading
Sample throughput	8/16/32	32
Fluorescence channels	3 / 4 / 5	4 / 5 / 6 / 7
Droplets per reaction	20,000	20,000
Multiplexing	Up to 18 targets per well	Up to 26 targets per well
Precision	±10%	±10%
Automation	Scalable: add generation/PCR units as volume grows; one reader serves several units	Fully automated after cartridge loading
Certification	CE-IVDR, FDA 510(k), NMPA	CE-IVDR
Best for	High-throughput labs: several units run samples in parallel, loaded at different times, no need to pool samples; capacity grows easily	Laboratories seeking a single-instrument, walk-away workflow

* Assay sensitivity is not a single instrument specification: it depends on target, input copies, specimen, workflow and validated detection rules. No universal assay LoD is claimed here.

* Time to result is not a universal claim. It depends on specific thermal cycling protocols.

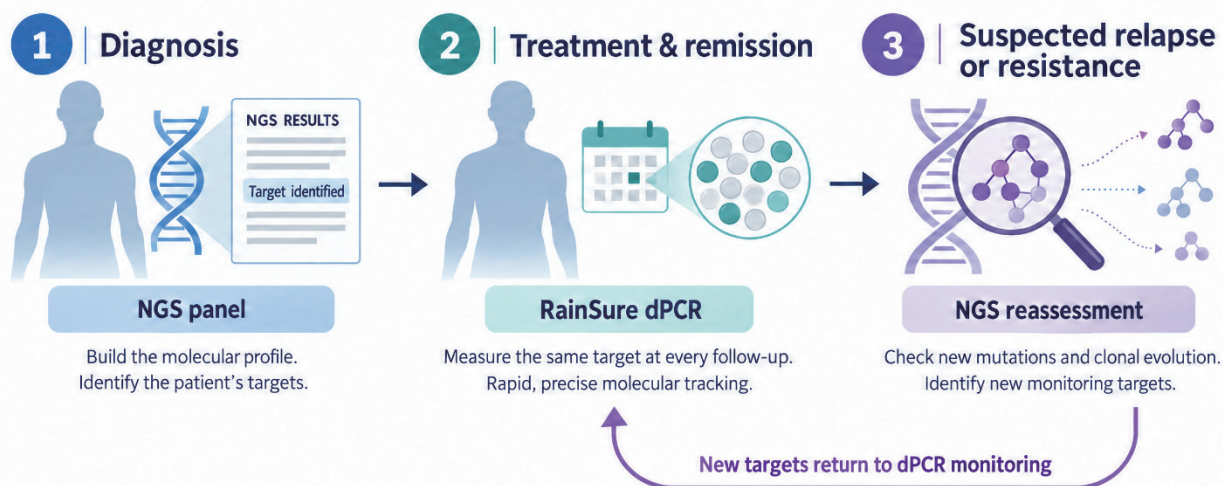
Why NGS + dPCR

NGS finds the target. dPCR follows it.

In hematologic malignancies, each patient carries a specific set of mutations and fusion genes. NGS gives the complete picture at diagnosis: it screens many genes at once and identifies which molecular targets are worth following. From that moment on, the question changes: no longer "what is there?", but "how much is left, and is it rising?". This is where digital PCR comes in. Once the target is known, RainSure dPCR measures it precisely and repeatedly, at every follow-up, quickly and at low cost. If the disease changes, NGS is used again to look for new mutations, and the new targets go back to dPCR.

THE PATIENT JOURNEY

Discover with NGS. Track with dPCR. Reassess as disease evolves.



Molecular coverage is panel-dependent. Monitoring requires suitable targets and validated dPCR assays.

Why dPCR for follow-up

- **High sensitivity.** RainSure dPCR detects the target down to 0.1%-0.001%, to follow low-level residual disease.
- **Absolute quantification.** Copies are counted directly, without a standard curve, so results can be compared from one time point to the next.
- **Sustainable over time.** Testing one known target costs far less than repeating a full NGS panel at every follow-up.
- **Earlier warning.** A rising molecular signal can appear before morphological relapse, giving the clinical team time to act (see Case 2).

Case Studies

CASE 1

Use of digital PCR for quantitative detection of genetic mutations in post-transplant patients

AML after transplant · patient-specific TP53 and ATM monitoring

Clinical question

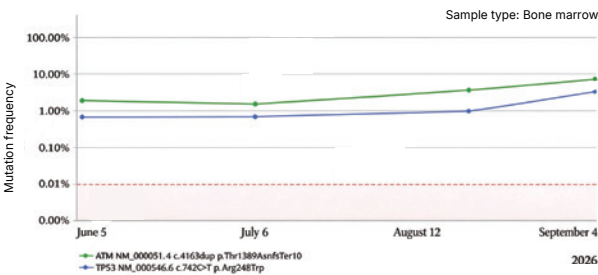
After transplant, are the patient's previously identified mutations persisting or rising?

What NGS found

On 19 December 2025, a 74-gene myeloid panel identified TP53 c.742C>T (p.Arg248Trp; VAF 37.8%) and ATM c.4163dup (reported VAF 70.16%). Screening for 18 common AML fusion genes by RT-PCR was negative. Both mutation rows refer to the myeloid gene-mutation test.

What dPCR measured

Target	June 2026	September 2026	Sample
TP53 c.742C>T	0.68%	3.48%	Bone marrow
ATM c.4163dup	1.96%	7.39%	Bone marrow



$$\text{Mutation frequency} = \frac{\text{Mutant copies}}{\text{Mutant copies} + \text{Wild-type copies}} \times 100\%$$

Endpoint values from the review notes; lines connect the two supplied measurements only. No intermediate data or unverified 0.01% threshold is inferred.

Result and why it matters

Both tracked mutation signals rose between June and September 2026. In this patient, the pattern supports further assessment alongside bone-marrow morphology, flow-cytometry MRD and other clinical findings; it does not independently establish relapse.

ASSAY USED: Custom assays for TP53 c.742C>T and ATM c.4163dup were used in this study.

After transplant, RainSure dPCR tracked the patient's own mutations and flagged a rising trend for clinical review.

CASE 2

Digital PCR technology for post-treatment quantification of fusion genes

ALL · IKZF2::NOTCH1 fusion monitoring

Clinical question

Does a previously identified fusion signal reappear during follow-up?

What NGS found

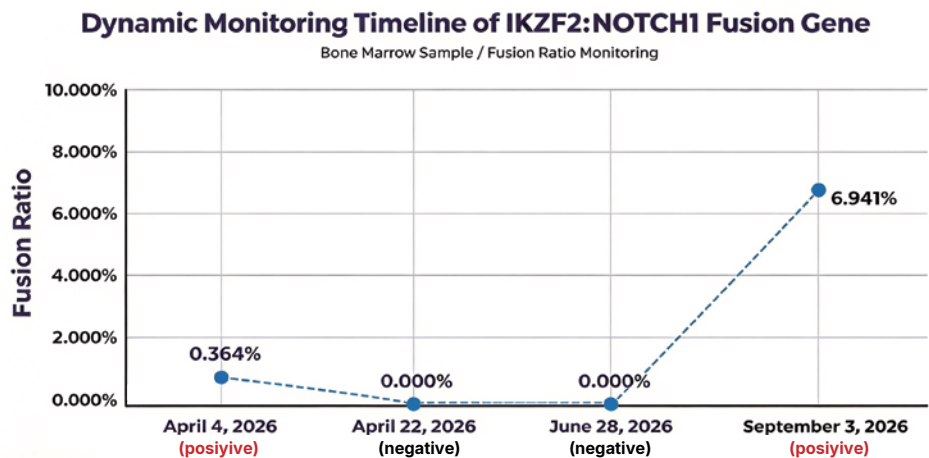
Whole-transcriptome sequencing on 13 March 2026 identified IKZF2::NOTCH1, with 112 supporting reads. The first dPCR measurement on 4 April was a follow-up measurement, not the diagnostic baseline.

What dPCR measured

Date (2026)	Sample	Fusion copies	Reference copies	Ratio / result
4 Apr	Bone marrow	130	35,760	0.364% · Positive
22 Apr	Bone marrow	0	31,560	Not detected*
28 Jun	Bone marrow	0	209,440	Not detected*
3 Sep	Bone marrow	6,880	99,120	6.941% · Positive

Ratio = fusion copies / reference copies × 100.

*No fusion copies were detected in these tested samples; this is not proof of biological absence



Result and why it matters

Two negative follow-up samples were followed by a positive 6.941% signal on 3 September. After communication between the Kindstar laboratory and the clinical team, recurrence was confirmed. The date of clinical confirmation was not supplied, so no lead time is claimed.

ASSAY USED Custom IKZF2::NOTCH1 fusion assay; not included in the supplied 48-assay fusion hotspot panel. Custom assay provided for this study.

Continuous test results: Positive, Negative, Negative, then Positive: RainSure dPCR flagged recurrence, later confirmed by the clinical team.

CASE 3

Digital PCR technology for fusion gene detection

B-ALL · IKZF1::IKZF1 and SSBP2::CSF1R monitoring

Clinical question

Do two patient-specific fusion signals track the same post-treatment pattern as immune-repertoire sequencing?

What NGS found

Whole-transcriptome sequencing on 3 March 2026 identified IKZF1::IKZF1 (13 supporting reads) and SSBP2::CSF1R (65 supporting reads). These are the source-reported fusion names; the IKZF1 transcript structure needs confirmation.

What dPCR measured

Date (2026)	Fusion	Fusion copies	Ref. copies	Ratio
9 Apr	IKZF1::IKZF1	944	127,600	0.740%
9 Apr	SSBP2::CSF1R	952	131,280	0.725%
28 May	IKZF1::IKZF1	18	81,000	0.022%
28 May	SSBP2::CSF1R	19	87,320	0.022%
7 Aug	IKZF1::IKZF1	20	145,200	0.014%
7 Aug	SSBP2::CSF1R	14	148,360	0.009%
22 Aug	IKZF1::IKZF1	10	99,560	0.010%
22 Aug	SSBP2::CSF1R	21	130,600	0.016%

All samples were bone marrow; all eight results were reported positive. Ratio = fusion copies / reference copies × 100.

Immune-repertoire sequencing

Date	IGH-SEQ1	IGH-SEQ2	IGK-SEQ3
3 Mar 2026	22.83718%	15.32433%	18.26181%
8 Apr 2026	0.18458%	0.13442%	1.01745%*
27 May 2026	0.00225%	0.00143%	0.00860%
6 Aug 2026	0.00945%	0.00383%	0.06050%
21 Aug 2026	0.00110%	0.00089%	0.01144%

Result and why it matters

Both fusion ratios fell markedly and remained detectable. Immune-repertoire sequencing also showed a substantial decline with residual positive signals. The methods are directionally consistent, but their sampling dates and percentage scales differ; exact agreement at every time point is not established.

ASSAY USED SSBP2::CSF1R: custom assay. IKZF1::IKZF1 custom assay used in this study.

Two fusion targets and an independent sequencing method showed the same overall pattern: a large decline with residual detectable signal.

What you can order

NGS defines the target. 167 RainSure dPCR loci monitor it.

167 dPCR loci for MRD monitoring

Fusion genes (46) Classic fusions incl. BCR::ABL1, PML::RARA, ETV6::RUNX1 and KMT2A / NUP98 rearrangements, across leukemia and lymphoma.

46 fusion-gene loci

Hotspot mutations (121) High-frequency drivers incl. FLT3-ITD/TKD, NPM1, JAK2 V617F, NRAS/KRAS, IDH1/2, TP53, ASXL1, SF3B1.

121 mutation loci across 43 genes

Not on the list? Match the NGS finding to a ready locus, or build a patient-specific custom assay with RainSure.

ORDERING INFORMATION

Product	Content	Catalog No
Hematology fusion-gene dPCR assays	46 fusion-gene loci, incl. BCR::ABL1, PML::RARA, ETV6::RUNX1	3.02.01.4440
Hematology mutation dPCR assays	121 hotspot mutation loci across 43 genes	3.02.01.4441
BCR ABL p210/p190 digital PCR one-tube quant test (CE-IVD)	Quantification of BCR-ABL1 p210 (e13a2/e14a2) and p190 (e1a2) fusion transcripts in human peripheral blood	3.02.03.0001
FastPlex™ BCR-ABL (p210) %IS digital PCR Kit – FDA 510(k) cleared	Quantification for monitoring major (e13a2/e14a2) BCR-ABL1 transcripts in CML patients	3.02.03.0009
Human JAK2 Gene V617F Mutation Detection Kit	Quantification of the JAK2 gene V617F mutation in DNA samples	3.02.03.0006
General Reagents	Digital PCR Mastermix for Probes	3.02.01.4066
	4×Digital PCR Mastermix for Probes	3.02.01.4388
	Lyophilized Digital PCR Mastermix for Probes	3.02.01.4380
	4× digital PCR Multiplex Mastermix for probes	3.02.01.4348
General Consumables	cartridges, cartridge cover and heat film	3.04.01.4001,3.04.01.4005;3.04.01.4003
Custom hematology dPCR assay service	Patient- or lab-specific targets: design, transfer, validation	

Ordering for 3.02.01.4440 and 3.02.01.4441: Each line in the catalog indicates the various test items; digital PCR assay products for different fusions/mutations can be ordered separately under the same catalog number according to the product serial number and name.

Custom assay enquiries

Provide the gene/fusion, full variant or transcript identifier, specimen and intended workflow. Confirm feasibility, analytical validation, reporting thresholds and quote details before ordering.

dPCR fusion gene locus assays (RUO)

Catalog No: 3.02.01.4440

All assays are provided in a 12-T package format (12 tests/tube). The assay is compatible with peripheral blood and bone marrow samples. The catalog includes 46 distinct kits.

Disease	No.	Fusion Gene	Detection Targets of Gene Fusions and Chromosomal Rearrangements Associated with Hematological Malignancies	Package Specification
Myeloid leukemia	1	BCR::ABL1 p210	Classic Philadelphia fusion; mainly CML; minority in Ph+ ALL	12 tests / tube
	2	BCR::ABL1 P230	Neutrophilic-predominant variant (e19a2)	12 tests / tube
	3	PML::RARA BCR1	t(15;17) long form (L-type); may co-occur with FLT3-ITD	12 tests / tube
	4	PML::RARA BCR3	t(15;17) short form (S-type); often with FLT3-ITD	12 tests / tube
	5	RUNX1::RUNX1T1	t(8;21); AML-M2; favorable prognosis	12 tests / tube
	6	CBFB::MYH11	inv(16); AML-M4Eo; favorable prognosis	12 tests / tube
	7	FUS::ERG	FUS-ERG; aggressive AML	12 tests / tube
	8	NUP98::NSD1	NUP98-NSD1; pediatric and normal-karyotype AML; poor prognosis	12 tests / tube
	9	NUP98::HOXA9	t(7;11); NUP98-HOXA9	12 tests / tube
	10	NUP98::KDM5A	NUP98-KDM5A	12 tests / tube
	11	NUP98::PRRX2	NUP98-PRRX2	12 tests / tube
	12	NUP98::TOP1	NUP98-TOP1	12 tests / tube
	13	NUP98::HOXD13	NUP98-HOXD13	12 tests / tube
	14	ETV6::MECOM	3q26 rearrangement; MECOM aberration	12 tests / tube
	15	KAT6A::CREBBP	t(8;16); monocytic leukemia	12 tests / tube
	16	CBFA2T3::GLIS2	CBFA2T3-GLIS2; infant AML; poor prognosis	12 tests / tube
	17	KMT2A::MLLT3(MLL/AF9)	t(9;11); predominantly AML-M5	12 tests / tube
	18	KMT2A::PICALM	KMT2A-PICALM	12 tests / tube
	19	KMT2A::MLLT10	t(10;11); KMT2A-MLLT10	12 tests / tube
	20	KMT2A::EPS15	KMT2A-EPS15	12 tests / tube
	21	KMT2A::MYO1F	KMT2A-MYO1F	12 tests / tube
	22	NPM1::MLF1	t(3;5); NPM1-MLF1	12 tests / tube
	23	RUNX1::CBFA2T3	RUNX1-CBFA2T3; rare	12 tests / tube
	24	RUNX1::MECOM	3q26 rearrangement; RUNX1-MECOM	12 tests / tube
	25	BCR::FGFR1	8p11 rearrangement; may present as MPN/AML/T-ALL	12 tests / tube
	26	HNRNPH1::ERG	HNRNPH1-ERG	12 tests / tube
	27	RBM15::MKL1	t(1;22); acute megakaryoblastic leukemia	12 tests / tube
	28	ZBTB16::RARA	t(11;17) variant APL; resistant to ATRA/arsenic trioxide	12 tests / tube
Acute lymphoblastic leukemia	29	BCR::ABL1 p190	Main fusion in Ph+ ALL (e1a2); occasionally in CML	12 tests / tube
	30	ETV6::RUNX1	t(12;21); most common in pediatric B-ALL; favorable prognosis	12 tests / tube
	31	KMT2A::AFF1(MLL-AF4)	t(4;11); infant and high-risk B-ALL	12 tests / tube

Disease	No.	Fusion Gene	Detection Targets of Gene Fusions and Chromosomal Rearrangements Associated with Hematological Malignancies	Package Specification
T/NK-cell lymphoma	32	TCF3::PBX1	t(1;19); pre-B ALL	12 tests / tube
	33	STIL::TAL1	STIL-TAL1 microdeletion; common molecular aberration in T-ALL	12 tests / tube
	34	EBF1::PDGFRB	t(5;12); eosinophilia-associated; imatinib-sensitive	12 tests / tube
	35	TCF3::ZNF384	ZNF384-rearranged B-ALL (WHO-HAEM5 entity)	12 tests / tube
	36	MEF2D::HNRNPUL1	MEF2D-rearranged B-ALL (new entity); poor prognosis	12 tests / tube
	37	TAF15::ZNF384	ZNF384-rearranged B-ALL (new entity)	12 tests / tube
	38	KMT2A::MLLT1	t(19;11); KMT2A-MLLT1	12 tests / tube
	39	ARID1B::ZNF384	ZNF384-rearranged B-ALL (new entity)	12 tests / tube
	40	NUP214::ABL1	ABL1 kinase domain fusion in T-ALL (TKI-sensitive)	12 tests / tube
	41	SET::NUP214	SET-NUP214; rare fusion in T-ALL	12 tests / tube
	42	PAX5::CBFA2T3	PAX5-rearranged B-ALL	12 tests / tube
	43	CREBBP::ZNF384	ZNF384-rearranged B-ALL (new entity)	12 tests / tube
	44	EP300::ZNF384	ZNF384-rearranged B-ALL (new entity)	12 tests / tube
	45	MEF2D::BCL9	MEF2D-rearranged B-ALL (new entity)	12 tests / tube
	46	NPM1::ALK	t(2;5); ALK+ ALCL	12 tests / tube

dPCR gene-mutation locus assays (RUO)

Catalog No: 3.02.01.4441

All assays are provided in a 12-T package format (12 tests/tube). The assay is compatible with peripheral blood and bone marrow samples. The catalog includes 121 distinct kits.

Disease	No.	Mutation	Common designation	Package Specification
Myeloid leukemia	1	c.35G>A p.Gly12Asp	NRAS G12D	12 tests / tube
	2	c.38G>A p.Gly13Asp	NRAS G13D	12 tests / tube
	3	c.34G>A p.Gly12Ser	NRAS G12S	12 tests / tube
	4	c.182A>G p.Gln61Arg	NRAS Q61R	12 tests / tube
	5	c.34G>T p.Gly12Cys	NRAS G12C	12 tests / tube
	6	c.181C>A p.Gln61Lys	NRAS Q61K	12 tests / tube
	7	c.38G>T p.Gly13Val	NRAS G13V	12 tests / tube
	8	c.35G>C p.Gly12Ala	NRAS G12A	12 tests / tube
	9	c.37G>T p.Gly13Cys	NRAS G13C	12 tests / tube
	10	c.37G>C p.Gly13Arg	NRAS G13R	12 tests / tube
	11	c.183A>T p.Gln61His	NRAS Q61H	12 tests / tube
	12	c.183A>C p.Gln61His	NRAS Q61H	12 tests / tube
	13	c.35G>T p.Gly12Val	NRAS G12V	12 tests / tube
	14	c.182A>T p.Gln61Leu	NRAS Q61L	12 tests / tube
	15	c.2503G>T p.Asp835Tyr	FLT3 D835Y	12 tests / tube
	16	c.2503G>C p.Asp835His	FLT3 D835H	12 tests / tube
	17	c.2028C>A p.Asn676Lys	FLT3 N676K	12 tests / tube
	18	c.2508_2510del p.Ile836del	FLT3 I836del	12 tests / tube

Disease	No.	Mutation	Common designation	Package Specification
Myeloid leukemia	19	c.2504A>T p.Asp835Val	FLT3 D835V	12 tests / tube
	20	c.2516A>G p.Asp839Gly	FLT3 D839G	12 tests / tube
	21	c.2505T>A p.Asp835Glu	FLT3 D835E	12 tests / tube
	22	c.2505T>G p.Asp835Glu	FLT3 D835E	12 tests / tube
	23	c.2039C>T p.Ala680Val	FLT3 A680V	12 tests / tube
	24	c.226G>A p.Glu76Lys	PTPN11 E76K	12 tests / tube
	25	c.215C>T p.Ala72Val	PTPN11 A72V	12 tests / tube
	26	c.226G>C p.Glu76Gln	PTPN11 E76Q	12 tests / tube
	27	c.1508G>T p.Gly503Val	PTPN11 G503V	12 tests / tube
	28	c.214G>A p.Ala72Thr	PTPN11 A72T	12 tests / tube
	29	c.181G>T p.Asp61Tyr	PTPN11 D61Y	12 tests / tube
	30	c.218C>T p.Thr73Ile	PTPN11 T73I	12 tests / tube
	31	c.182A>G p.Asp61Gly	PTPN11 D61G	12 tests / tube
	32	c.227A>G p.Glu76Gly	PTPN11 E76G	12 tests / tube
	33	c.35G>A p.Gly12Asp	KRAS G12D	12 tests / tube
	34	c.38G>A p.Gly13Asp	KRAS G13D	12 tests / tube
	35	c.35G>T p.Gly12Val	KRAS G12V	12 tests / tube
	36	c.34G>T p.Gly12Cys	KRAS G12C	12 tests / tube
	37	c.35G>C p.Gly12Ala	KRAS G12A	12 tests / tube
	38	c.34G>A p.Gly12Ser	KRAS G12S	12 tests / tube
	39	c.351A>T p.Lys117Asn	KRAS K117N	12 tests / tube
	40	c.437C>T p.Ala146Val	KRAS A146V	12 tests / tube
	41	c.2466T>G p.Asn822Lys	KIT N822K	12 tests / tube
	42	c.2466T>A p.Asn822Lys	KIT N822K	12 tests / tube
	43	c.1255_1257del p.Asp419del	KIT D419del	12 tests / tube
	44	c.2447A>T p.Asp816Val	KIT D816V	12 tests / tube
	45	c.2446G>T p.Asp816Tyr	KIT D816Y	12 tests / tube
	46	c.2446G>C p.Asp816His	KIT D816H	12 tests / tube
	47	c.2467T>G p.Tyr823Asp	KIT Y823D	12 tests / tube
	48	c.394C>T p.Arg132Cys	IDH1 R132C	12 tests / tube
	49	c.395G>A p.Arg132His	IDH1 R132H	12 tests / tube
	50	c.394C>G p.Arg132Gly	IDH1 R132G	12 tests / tube
	51	c.394C>A p.Arg132Ser	IDH1 R132S	12 tests / tube
	52	c.395G>T p.Arg132Leu	IDH1 R132L	12 tests / tube
	53	c.1934dup p.Gly646TrpfsTer12	ASXL1 G646Wfs*12	12 tests / tube
	54	c.2003dup p.Gly669TrpfsTer2	ASXL1 G669Wfs*2	12 tests / tube
	55	c.1900_1922del p.Glu635ArgfsTer15	ASXL1 E635Rfs*15	12 tests / tube
	56	c.2572C>T p.Gln858Ter	ASXL1 Q858*	12 tests / tube
	57	c.3202C>T p.Arg1068Ter	ASXL1 R1068*	12 tests / tube

Disease	No.	Mutation	Common designation	Package Specification
Myeloid leukemia	58	c.860_863dup p.Trp288CysfsTer12	NPM1 W288Cfs*12	12 tests / tube
	59	c.863_864insCATG p.Trp288CysfsTer12	NPM1 W288Cfs*12	12 tests / tube
	60	c.863_864insCCTG p.Trp288CysfsTer12	NPM1 W288Cfs*12	12 tests / tube
	61	c.863_864insCAGA p.Trp288CysfsTer12	NPM1 W288Cfs*12	12 tests / tube
	62	c.602G>A p.Arg201Gln	RUNX1 R201Q	12 tests / tube
	63	c.1264G>T p.Glu422Ter	RUNX1 E422*	12 tests / tube
	64	c.1315dup p.Ser439PhefsTer161	RUNX1 S439Ffs*161	12 tests / tube
	65	c.601C>T p.Arg201Ter	RUNX1 R201*	12 tests / tube
	66	c.419G>A p.Arg140Gln	IDH2 R140Q	12 tests / tube
	67	c.515G>A p.Arg172Lys	IDH2 R172K	12 tests / tube
	68	c.418C>T p.Arg140Trp	IDH2 R140W	12 tests / tube
	69	c.2645G>A p.Arg882His	DNMT3A R882H	12 tests / tube
	70	c.2644C>T p.Arg882Cys	DNMT3A R882C	12 tests / tube
	71	c.2363C>T p.Ala788Val	DNMT3A A788V	12 tests / tube
	72	c.101C>T p.Ser34Phe	U2AF1 S34F	12 tests / tube
	73	c.101C>A p.Ser34Tyr	U2AF1 S34Y	12 tests / tube
	74	c.104G>T p.Arg35Leu	U2AF1 R35L	12 tests / tube
	75	c.1099_1150del p.Leu367ThrfsTer46	CALR L367Tfs*46 (type 1)	12 tests / tube
	76	c.1154_1155insTTGTC p.Lys385fsTer47	CALR K385fs*47 (type 2)	12 tests / tube
	77	c.912_913insTTG p.Lys304_Gln305insLeu	CEBPA K304_Q305insL	12 tests / tube
	78	c.68dup p.His24AlafsTer84	CEBPA H24Afs*84	12 tests / tube
	79	c.3916C>T p.Arg1306Ter	NF1 R1306*	12 tests / tube
	80	c.4102_4110+23delinsAAGG	—	12 tests / tube
	81	c.2608G>A p.Gly870Ser	SETBP1 G870S	12 tests / tube
	82	c.2602G>A p.Asp868Asn	SETBP1 D868N	12 tests / tube
	83	c.1156_1159dup p.Ala387ValfsTer4	WT1 A387Vfs*4	12 tests / tube
	84	c.1157C>A p.Ser386Ter	WT1 S386*	12 tests / tube
	85	c.1036_1037insCG p.Tyr346fsTer	ETV6 Y346fs*	12 tests / tube
	86	c.1154_1163del p.Asn385fsTer	ETV6 N385fs*	12 tests / tube
	87	c.1849G>T p.Val617Phe	JAK2 V617F	12 tests / tube
	88	c.1853C>T p.Thr618Ile	CSF3R T618I	12 tests / tube
	89	c.2098A>G p.Lys700Glu	SF3B1 K700E	12 tests / tube
	90	c.284C>G p.Pro95Arg	SRSF2 P95R	12 tests / tube
	91	c.1114G>A p.Ala372Thr	GATA2 A372T	12 tests / tube
	92	c.1574G>A p.Arg525His	DDX41 R525H	12 tests / tube
	93	c.1148G>A p.Trp383Ter	KMT2C W383*	12 tests / tube
	94	c.5594C>G p.Ala1865Gly	TET2 A1865G	12 tests / tube
	95	c.3215A>G p.Gln1072Arg	ZEB2 Q1072R	12 tests / tube

Disease	No.	Mutation	Common designation	Package Specification
Acute lymphoblastic leukemia	96	Ikaros-6 (Ik6) exon 4-7 deletion (Δ 4-7)	IKZF1 Ik6	12 tests / tube
	97	Ikaros-10 (Ik10) exon 2-7 deletion (Δ 2-7)	IKZF1 Ik10	12 tests / tube
	98	c.472G>A p.Gly158Ser	IKZF1 G158S	12 tests / tube
	99	c.2347C>T p.Leu783Phe	JAK1 L783F	12 tests / tube
	100	c.2878C>T p.Pro960Ser	JAK1 P960S	12 tests / tube
	101	c.737C>A p.Pro246Gln	PTEN P246Q	12 tests / tube
	102	c.1960C>T p.Arg654Ter	SUZ12 R654*	12 tests / tube
	103	c.2065C>T p.Arg689Trp	FBXW7 R689W	12 tests / tube
	104	c.346G>A p.Gly116Ser	GNB1 G116S	12 tests / tube
	105	c.955C>T p.Arg319Ter	PHF6 R319*	12 tests / tube
Chronic lymphocytic leukemia	106	c.524G>A p.Arg175His	TP53 R175H	12 tests / tube
	107	c.841G>C p.Asp281His	TP53 D281H	12 tests / tube
	108	c.733G>A p.Gly245Ser	TP53 G245S	12 tests / tube
	109	c.578A>G p.His193Arg	TP53 H193R	12 tests / tube
	110	c.817C>T p.Arg273Cys	TP53 R273C	12 tests / tube
	111	c.830G>A p.Cys277Tyr	TP53 C277Y	12 tests / tube
	112	c.5033T>C p.Leu1678Pro	NOTCH1 L1678P	12 tests / tube
	113	c.4793G>C p.Arg1598Pro	NOTCH1 R1598P	12 tests / tube
B-cell lymphoma	114	c.794T>C p.Leu265Pro	MYD88 L265P	12 tests / tube
	115	c.1799T>A p.Val600Glu	BRAF V600E	12 tests / tube
	116	c.4337G>A p.Arg1446His	CREBBP R1446H	12 tests / tube
	117	c.3826C>T p.Arg1276Ter	ARID1A R1276*	12 tests / tube
T/NK-cell lymphoma	118	c.1970G>A p.Arg657Gln	JAK3 R657Q	12 tests / tube
	119	c.1718C>T p.Ala573Val	JAK3 A573V	12 tests / tube
	120	c.1924A>C p.Asn642His	STAT5B N642H	12 tests / tube
	121	c.3295G>A p.Glu1099Lys	WHSC1 E1099K	12 tests / tube

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